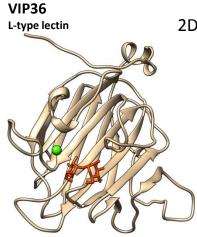
## **Critical Issues: Aspects of VIP36 Carbohydrate Interactions**

**Klaus Fiedler** 

### Roles of lectins: How can bound carbohydrates best be studied?



2DUR-A

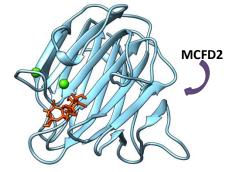
VIP36 stimulates secretion of clusterin Hara-Kuge et al. (2002)

VIP36 stimulates  $\alpha$ -amylase secretion Hara-Kuge et al. (2004)

the pH-dependence is controversial or small relative to VIPL or ERGIC53

ERGIC53 L-type lectin

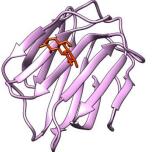
4GKX-A



MCFD2 binds coagulation factors V and VIII; some proteins bind directly to the lectin Kamiya et al. (2008)

Galectin-9 Galectin

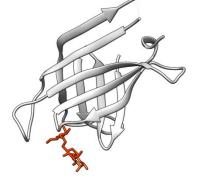
3LSE-A



Recognizes internal N-acetyllactosamine units <sub>Nagae et al. (2009)</sub>

#### OS-9

P-type lectin (mannose-6-phosphatereceptor homology domain)

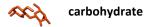


3AIH-A

Binds M8-M5 high mannose glycans via α6 / D3 / C arm tri-

Mannose Hosokawa et al. (2009) Satoh et al. (2010)

calcium

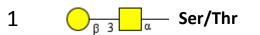


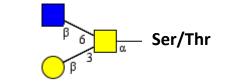
### Carbohydrates

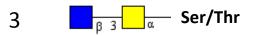
2

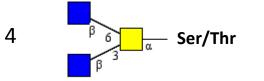
O Gal □ GalNAc ● Man O Glc □ GlcNAc ▲ Fuc ◆ NeuAc

O-glycan core

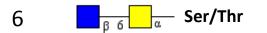


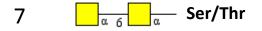


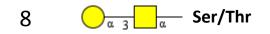


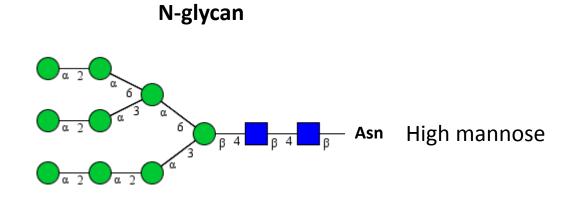


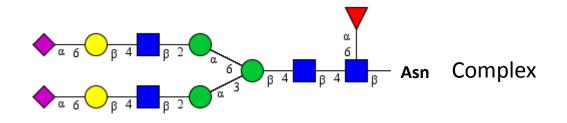
5 Garage Ser/Thr

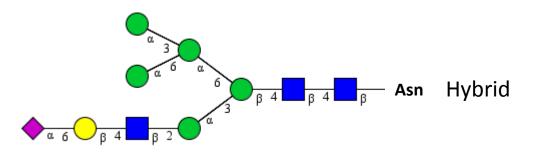












# Carbohydrate binding: 2DUR-A

Similarity of VIP36 to plant lectins was first described based on primary structure relative to the lectin from *Bauhinia* Fiedler et al. (1994)

The *Bauhinia* lectin was yet not retrieved in this search

#### Structural Search Programme PDBeFold

Protein	Structure	Q-Score	RMSD	Species
VIP36	2dur	1	0.00	Canis lupus familiaris
ERGIC53	4gkx	0.7	1.08	Homo sapiens
EMP47	2a71	0.52	1.80	Saccharomyces cerevisiae
EMP46	2a6w	0.48	1.95	Saccharomyces cerevisiae
Pea Lectin	2bqp	0.43	2.01	Pisum sativum
Soybean Agglutinin	2sba	0.41	1.88	Glycine max
Basic Agglutinin	2zml	0.41	1.97	Psophocarpus tetragonolobus
Seed Lectin	4u2a	0.40	2.07	Vatairea macrocarpa
solectin B4	1n47	0.40	1.94	Vicia villosa
Lectin DB58	1lul	0.40	1.92	Vigna unguiculata
Lectin	2eig	0.40	2.21	Lotus tetragonolobus
yesU	1oq1	0.33	2.39	Prokaryotic, Bacillus subtilis
Galectin-9	3lse	0.31	2.41	Homo sapiens
Galectin-7	5gal	0.31	2.07	Homo sapiens
Galectin-8	3ap4	0.30	2.31	Homo sapiens
Galectin-3	3zsl	0.30	2.26	Homo sapiens
Galectin-4	3i8t	0.29	2.44	Mus musculus
Galectin-1*	4no4	0.29	2.51	Rattus norvegicus
Galectin LEC-6	3vv1	0.29	2.24	Caenorhabditis elegans
Galectin-1	1sla	0.29	2.53	Bos taurus
Galectin-2	2ymz	0.29	2.52	Gallus gallus
Galectin-10 Charcot-Leyden	1g86	0.28	2.49	Homo sapiens
Calreticulin	300w	0.26	2.63	Mus musculus
Sialidase	2w68	0.24	2.33	Prokaryotic, Vibrio cholerae
Calnexin	1jhn	0.15	2.99	Canis lupus familiaris
VP4	1kqr	0.13	3.98	Virus, Rhesus rotavirus
Concanavalin A	1vam	0.12	1.91	Canavalia ensiformis
COMP	3fby	0.11	2.62	Homo sapiens

Krissinel

### **Protein glycosylation: Fucosylation**

N-glycan chains are modified by adding fucose residues to their terminal chains (fucosyltransferase 2; Fut2) or to the N-glycan core (fucosyltransferase 8; Fut8)

Fut2 activity has a protective role and was suggested to alter host-microbial interactions by terminal modifications (Pickard et al. 2014; Goto et al. 2014)

Core fucosylation by Fut8 has been described as possible sorting signal in protein traffic in hepatocytes to the bile duct (apical traffic)(Nakagawa et al. 2006)

HNF1 $\alpha$  is involved in the regulation of fucosylation (Lauc et al. 2010)

Regulatory role of IKZF1 in GlcNAc bisection versus core-fucosylation (Lauc et al. 2013)

### **Changes in protein glycosylation**

#### ... in Cancer

Fut8 is upregulated in epithelial-mesenchymal transition and was, furthermore, shown to regulate metastasis in nonsmall cell lung cancer (NSCLC) (Chen et al. 2013)

In breast, colorectal and ovarian cancer Fut8 is upregulated (Christiansen et al. 2014)

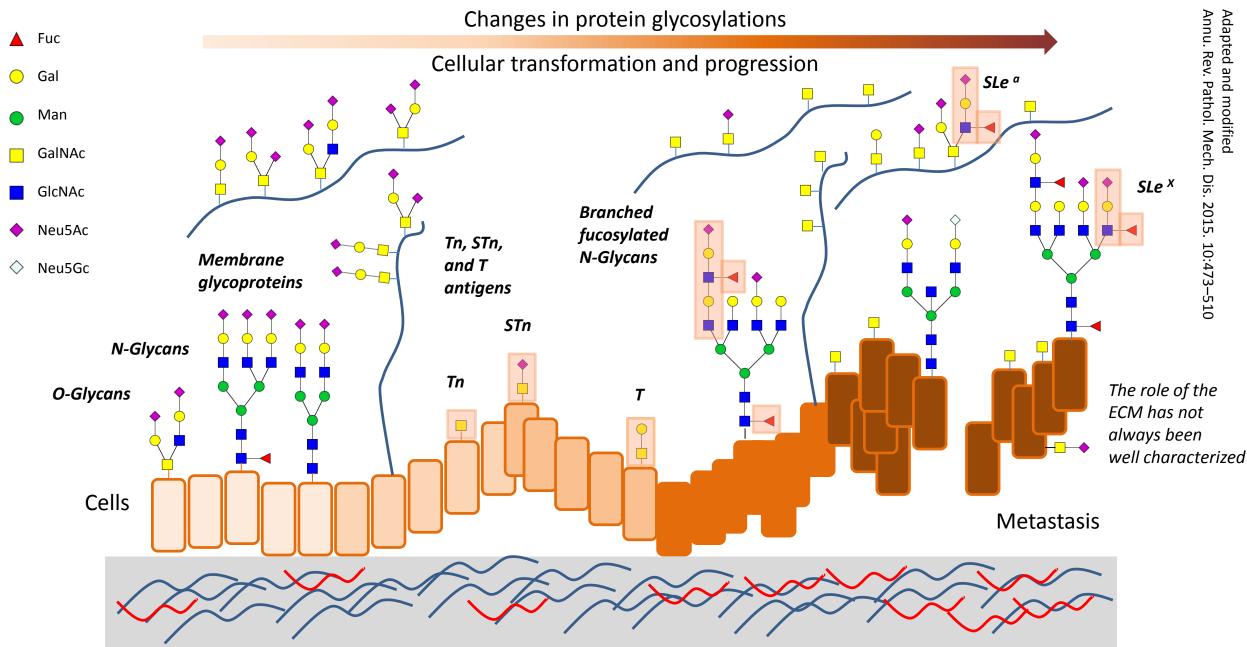
Fut8 overexpression in hepatoma cells (Hep3B) was demonstrated to suppress, however, intrahepatic metastasis in athymic mice (Miyoshi et al. 1999)

Absence of core fucose up-regulates GnT-III in mouse embryonic fibroblasts (Kurimoto et al. 2014)

In melanomas expression of fucokinase attenuates growth (Lau et al. 2015) (these results refer to both types of fucosylation)

First detailed Fut8 suppression in a systems approach shows the requirement for melanoma metastasis (Agrawal et al. 2017)

### Transformation in cells and glycosylation



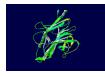
### VIP36 cellular localizations and early tissue data

Although evidence has been obtained for certain localization with the GFP (green fluorescent protein)tagging approach (Stadler et al. 2013) for proteins such as e.g. caveolin-1/VIP21 it has been shown, that these lead to aberrant localization (Han et al. 2015; Tiwari et al. 2016) and tagged constructs do not incorporate in natural endogenous complexes of proteins.

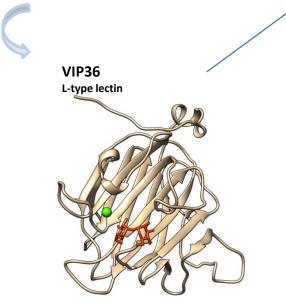
VIP36 has been analyzed in several cell types, endogenous VIP36 was found at the plasma membrane, the Golgi apparatus and intermediate compartment, and can be discerned in the Golgi apparatus in tissue sections (Human Protein Atlas; Uhlén et al. 2015). GFP-tagging approaches have failed on VIP36 and did not yield results consistent with the localization of the non-tagged protein obtained by raising antibodies against the extracellular domain or against natural peptides (Fiedler et al. 1994; Fiedler and Simons 1995; Füllekrug et al. 1999; Shimada et al. 2003). N- or C-terminal tagging approaches differed in resulting localization.

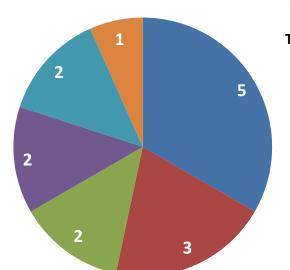
### VIP36 structure: Carbohydrate binding

VIP36 simulation (pre-structure data could be compared)



VIP36 binds to 2 calcium-ions when tested and simulated



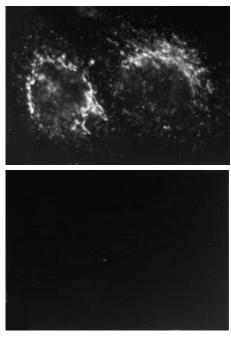


#### VIP36

Top scoring library glycans

- Lactosamine motif
- O-glycan
- N-glycan complex
- N-glycan high mannose
- N-glycan hybrid
- GSL

#### VIP36 localization



Fiedler and Simons (1995)

MCS search

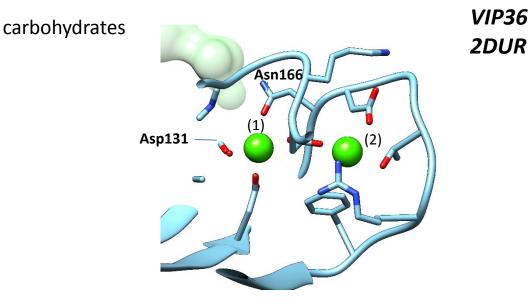
http://www.glycome-db.org

2DUR

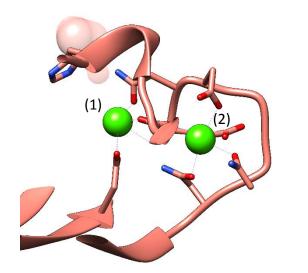
### VIP36 structure: Calcium binding retest in simulation

Previous data showed, that VIP36 binds to "1.8" calcium ions when tested in equilibrium dialysis and Scatchard analysis.

Calcium-independent affinity to carbohydrates has yet not been explained, the structural analysis, however, displayed calcium binding site (1) that is very close to the bound carbohydrate, and a distant cryptic calcium binding site (2). We had not found any dependence on manganese in the simple dialysis that would have been expected based on the similarity to leguminous lectins (Fiedler and Simons, 1994, 1995).



carbohydrates



ERGIC53 3WHU

> VIP36 binding to carbohydrates is affected by Ca<sup>2+</sup>, Ca<sup>2+</sup> binding itself may be experimentally affected by pH and the variable conditions used of pH 6.5 (crystal) versus 7.4 (solution) and associated alteration of partial charges of the aspartate groups (~0.4e unit charge).

### **Carbohydrate library**

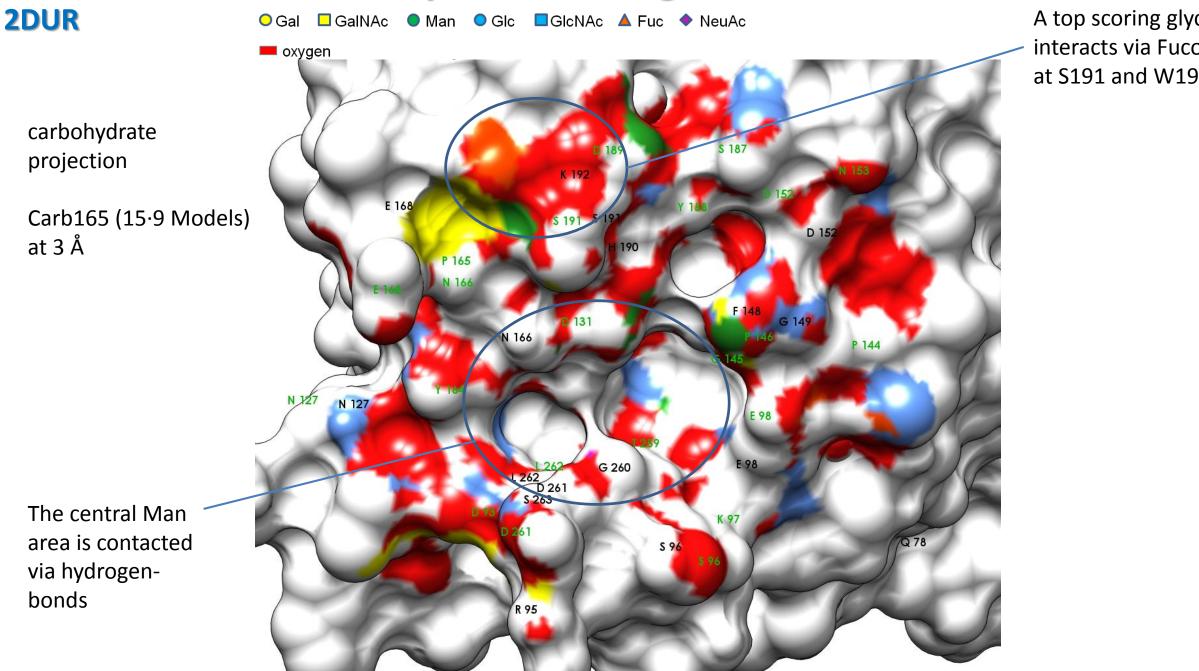


#### Library of Carbohydrates

N-glycan high mannose	19
N-glycan hybrid	17
N-glycan complex	9
Sialoside	27
Fucoside	25
O-glycan core	17
Other	51

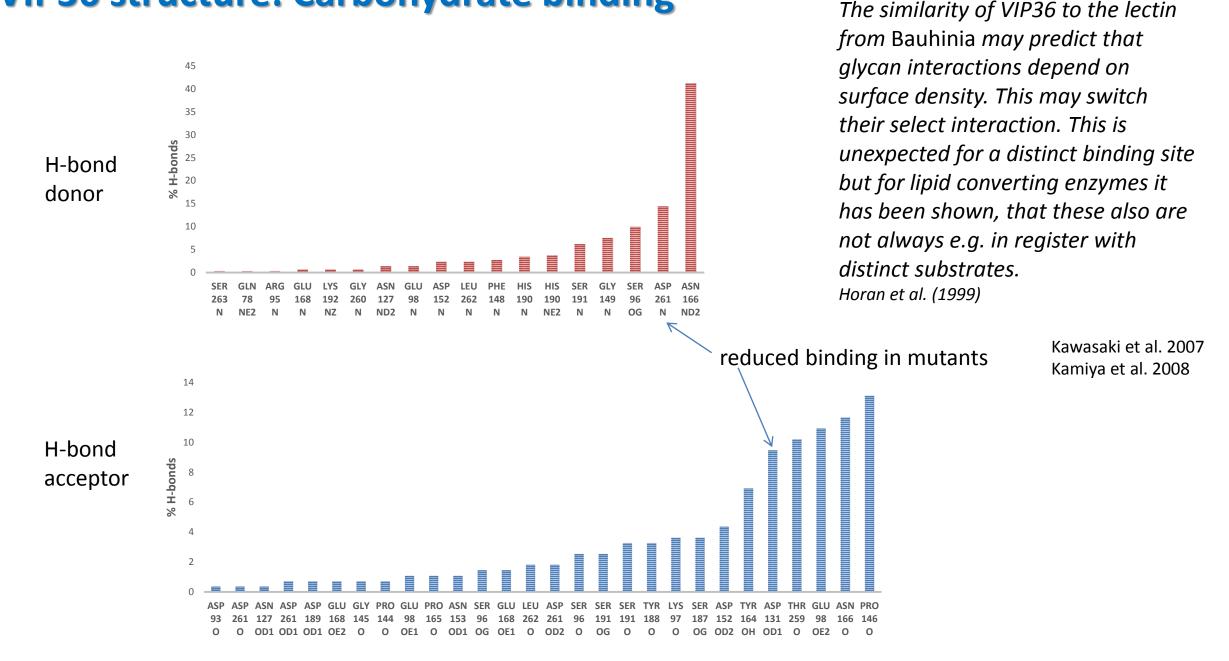
The library was assembled from internet sources: GLYCAM (http://www.ccrc.uga.edu) and glycoSCIENCES.DE (www.glycosciences.de)

### VIP36 structure: Carbohydrate binding

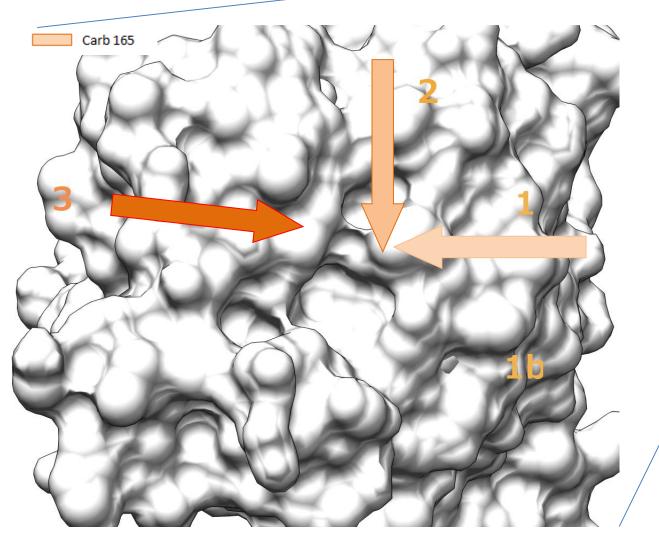


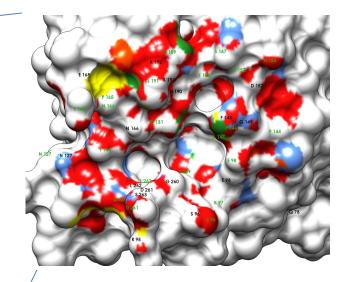
A top scoring glycan interacts via Fucose at \$191 and W196

### VIP36 structure: Carbohydrate binding



### **Carbohydrate binding: Topology**

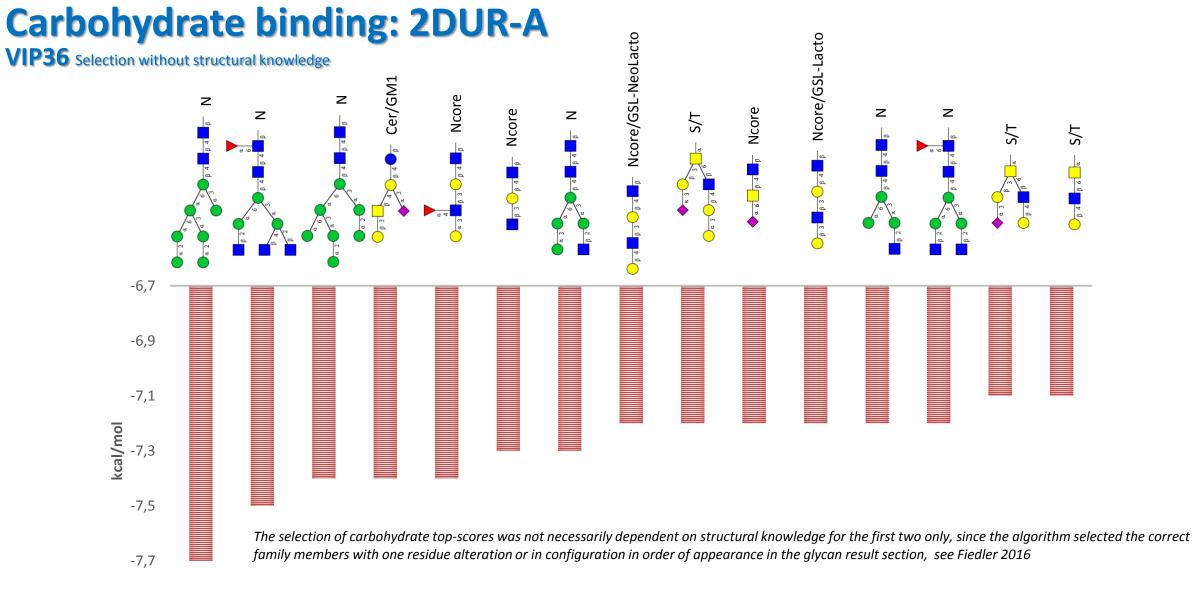




#### Topologies combining 15 top-scores of Carb165

Disulfide (C202-C239) distant from binding patch was not closed

Examples of glycan analogues in switched topology for some lectins are known (axial rotations)



Glc

Man

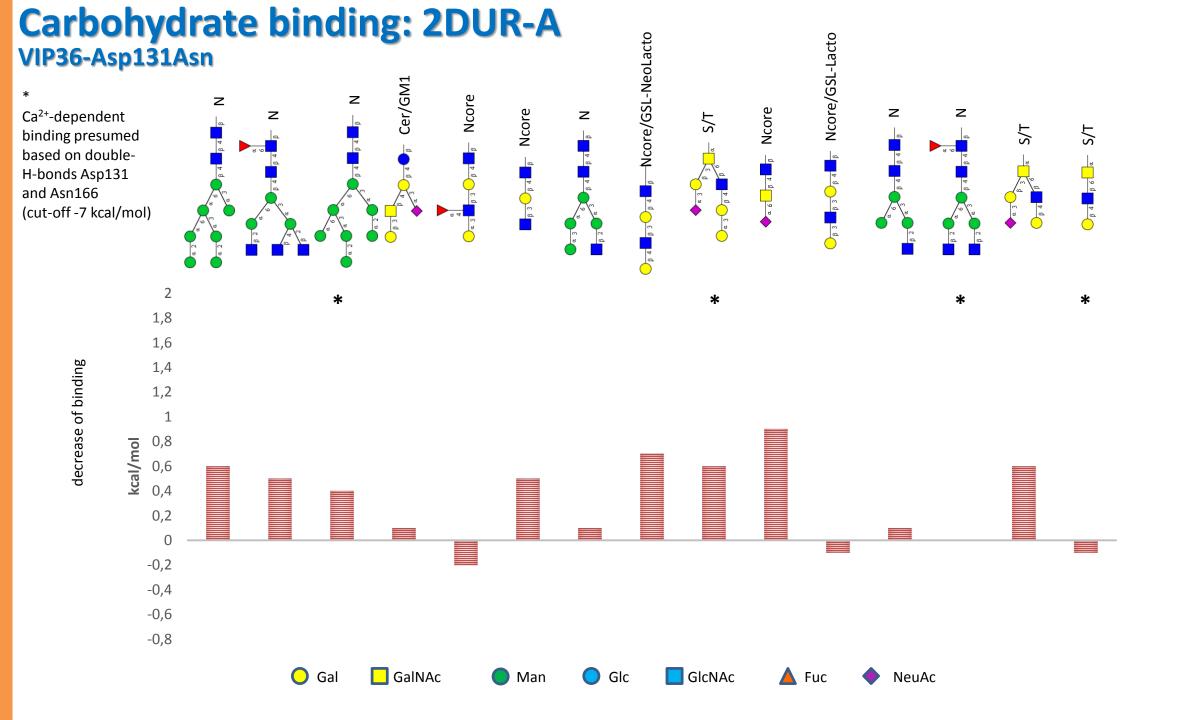
GlcNAc

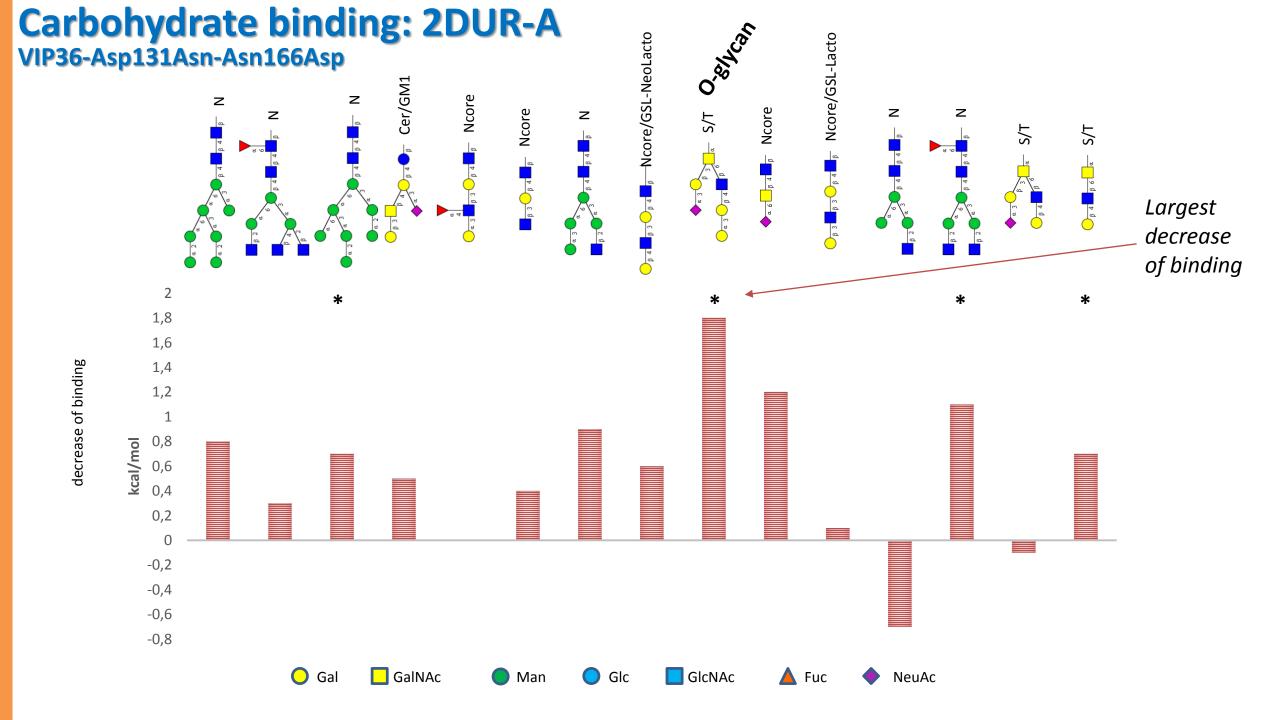
🛕 Fuc

NeuAc

-7,9

🔵 Gal 🛛 🗖 GalNAc

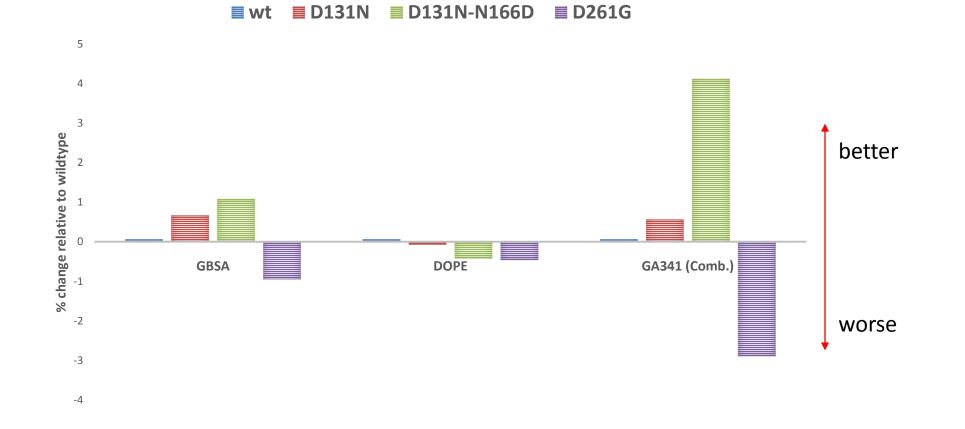




### **Carbohydrate binding: Mutant molecular folding functions**

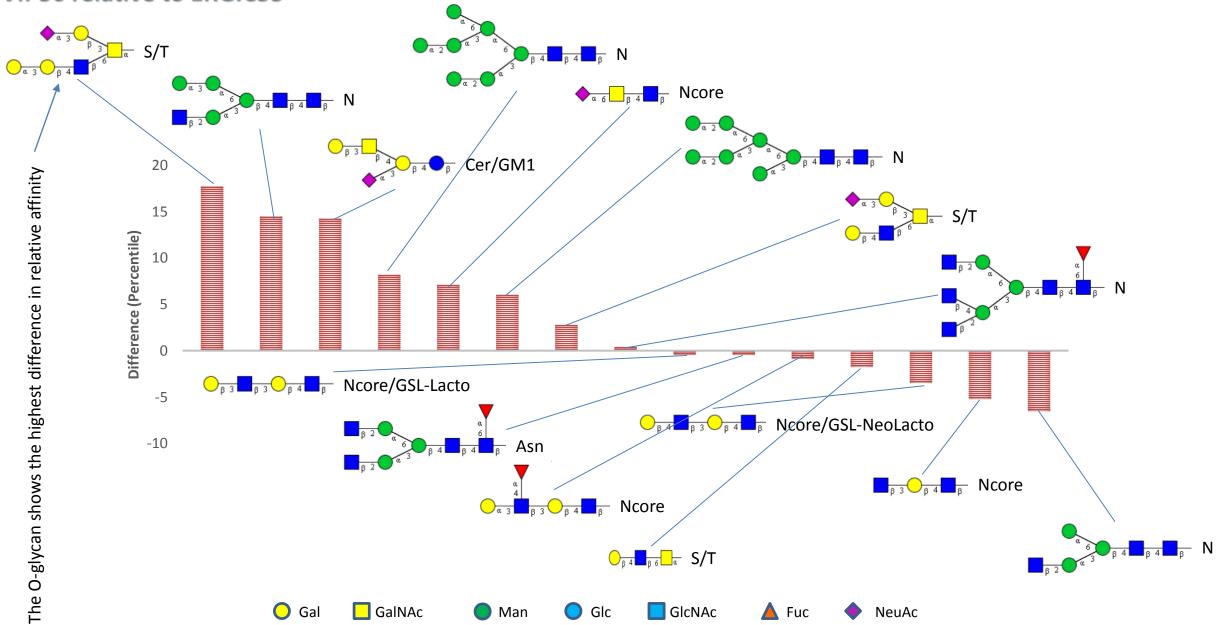
Scores from Fiser and Sali 2003, Shen and Sali 2006

See the binding site (colored figure) for an overview of the carbohydrate interaction

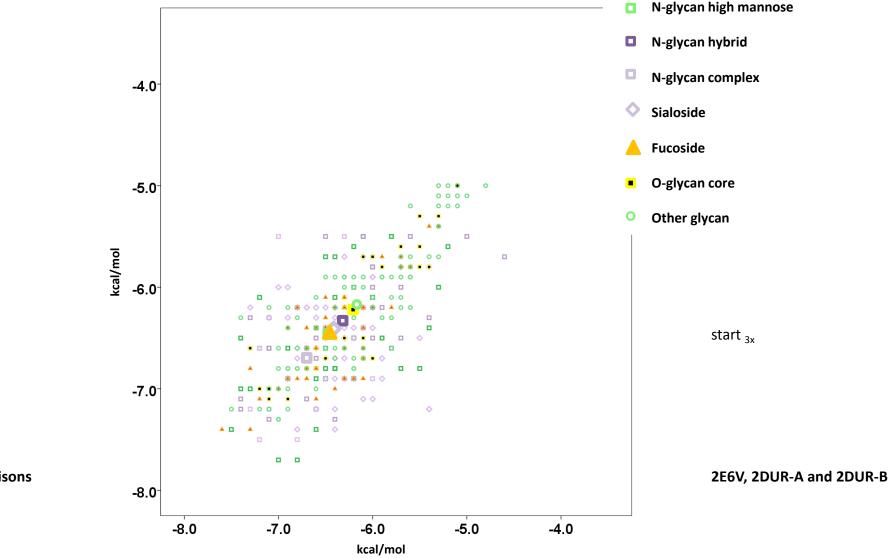


The DOPE score indicates good overall folding (D261G shows enhanced change) Scores determined after energy minimization

#### Carbohydrate binding: wt2DUR-A and wt4GKX-A VIP36 relative to ERGIC53

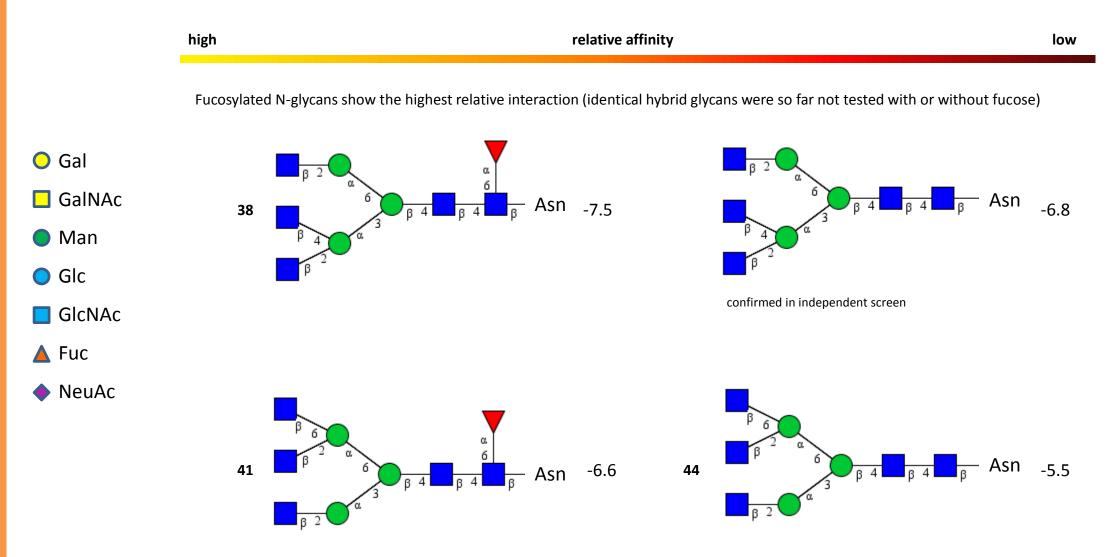


#### Carbohydrate binding: Summary VIP36 Own library data



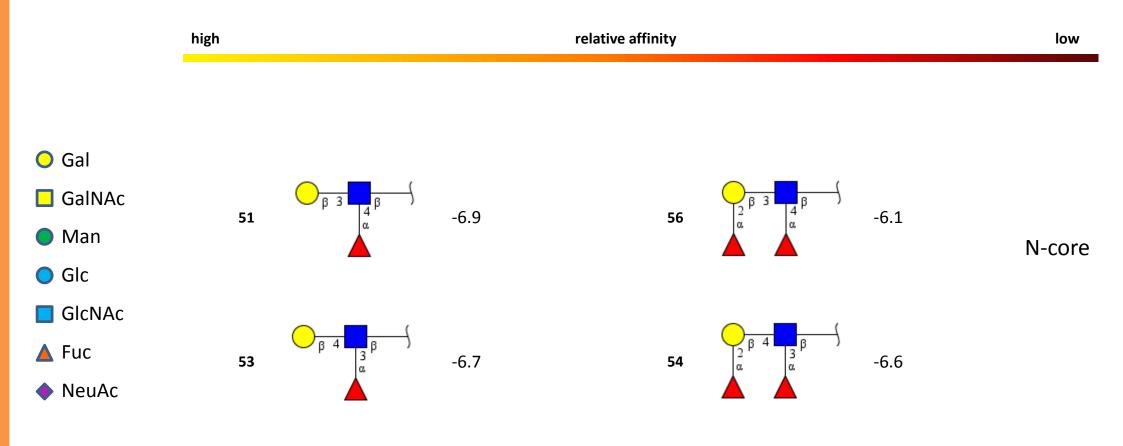
pairwise comparisons

### Carbohydrates interacting with VIP36: Top scores



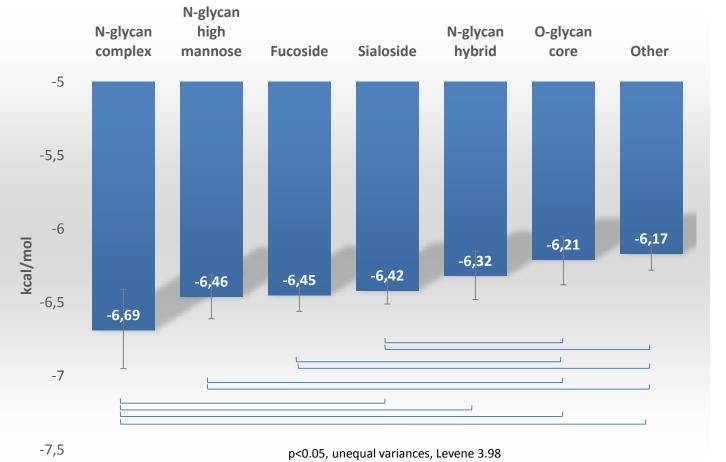
The  $\alpha$  1-6 linked fucose increases binding affinity by  $\leq$  -0.7 kcal/mol

### Carbohydrates interacting with VIP36: Top scores



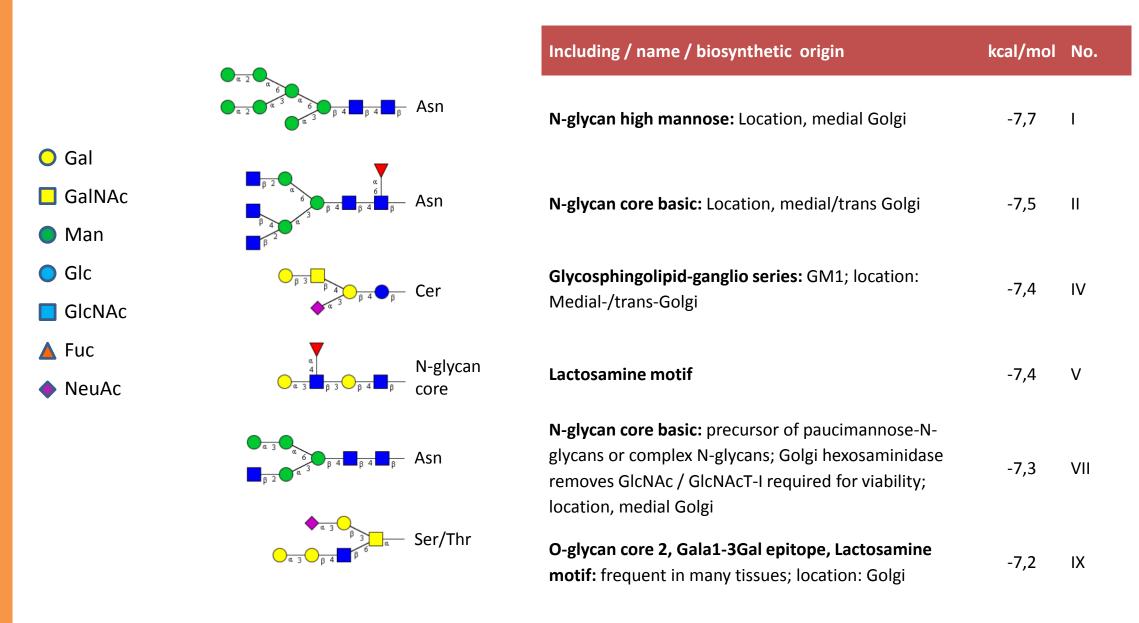
Other single fucoses do not increase binding affinity

#### Carbohydrate binding: Summary of VIP36 binding Group differences, Library data



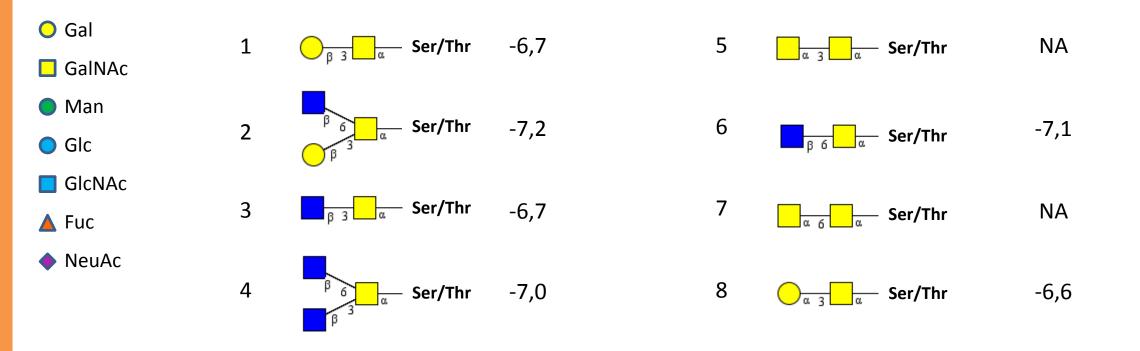
Confidence interval 95 %

### **Carbohydrate interactions: Top scores**



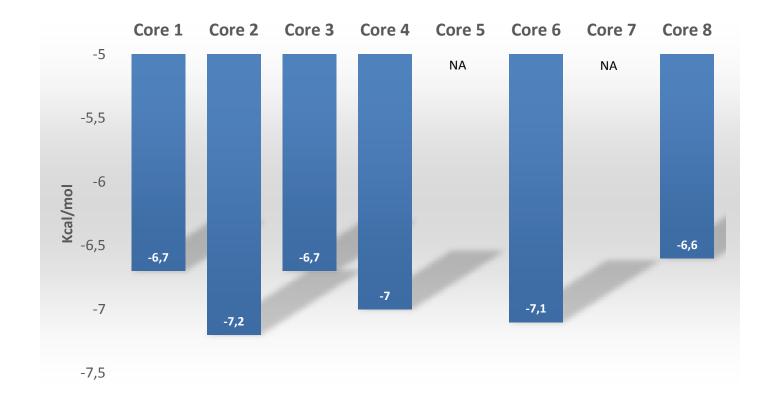
### **Carbohydrates interacting with VIP36: Top scores**

#### **O-glycan core including extension**



core 2 O-glycan only, and T antigen show lower binding affinity

#### Carbohydrates interacting with VIP36: Top scores O-glycan core including extension, Library data



### Fut8 in mice knockout strains

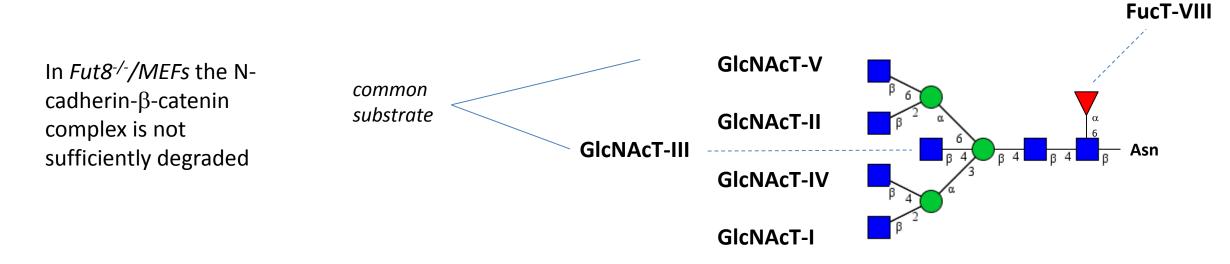
Fut8 knockout (Fut8<sup>-/-</sup>) mice show 70% mortality in the first 3 post-natal days

In surviving mice VEGFR-2 ist downregulated and the mice show emphysema-like changes of the lung

TGF- $\beta$ 1 receptor, EGF receptor and integrin activation show dysregulation

Wang et al. (2005, 2009)

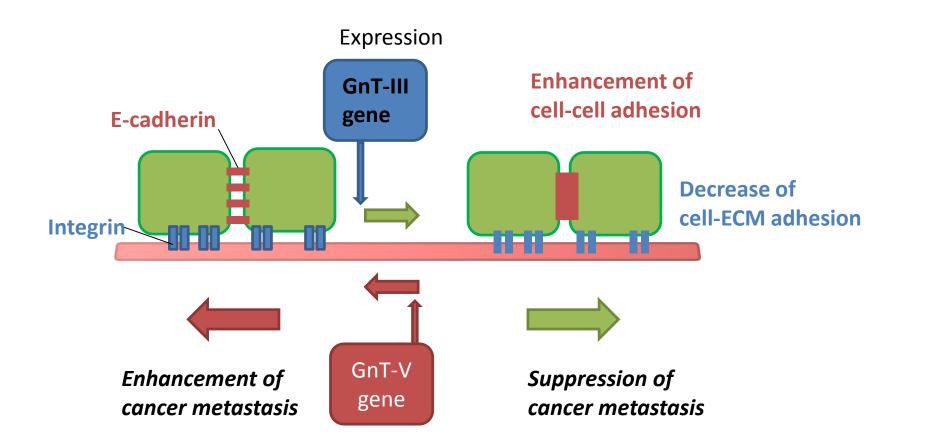
### The metastatic machinery in glycosylation?



Wnt target	Fut8 KO/WT x change (microarray)
Growth arrest-specific 1 (GAS1)	12.1
Frizzled homolog 6 (FZD6)	2.3
Glutathione S-transferase, type1 (GSTM1)	11.3
Cytochrome P450, CYP1B1	9.8
Mitochondrial aldehyde dehydrogenase 2 (ALDH2)	2.8
GlcNAcT-III	~3.0 (PCR)
GlcNAcT-V	0.3 (PCR)

Wnt-signal inhibitor (IWP-2) studies suggest that changes in Wnt/ β-catenin signalling are associated with the upregulation of GlcNAcT-III

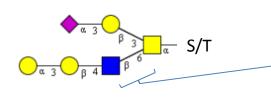
### Several roles of glycosyl-transferases in cancer



Adapted from Zhao et al. (2008)

According to this view, Fut8 would also indirectly affect metastasis via regulation of GlcNAc-transferases

### Several roles of glycosyl-transferases in cancer



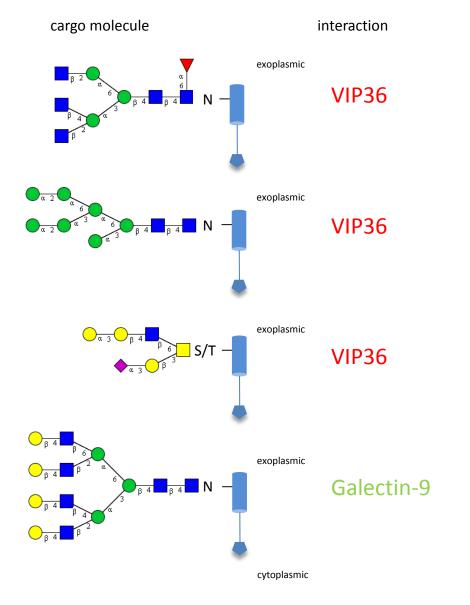
Core 2 O-glycans allow tumorous cells to evade Natural-Killer (NK) cells Benson et al. (2010)

Kidney tubule cell specific expression of core 2 GlcNAc-transferase

Suzuki et al. (2004)

May mucin transport in intestinal epithelia (core 3) and possibly kidney tubule specific O-glycans relate to VIP36 function?

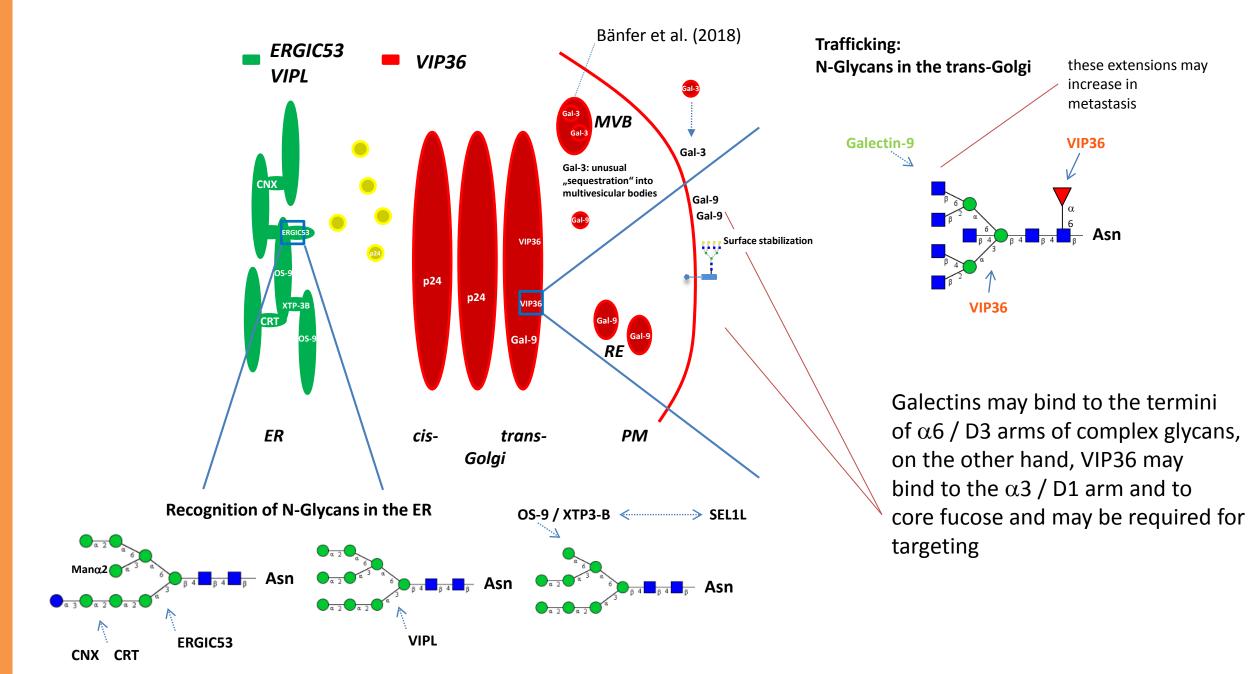
# Multiple possibilities of sorting/inclusion into transport carriers in the constitutive secretory pathway



- Regulated inclusion of cargo-receptors by a cytoplasmic coat
- Sorting in the Golgi lumen by domain formation and luminal matrix
- Cargo sequestering by substrate enzymatic conversions and release
- Surface/domain formation by stabilization

The capacities of a sorting machinery cannot be predicted off hand, therefore, the inverted Ushaped, or other dependence on cargo concentration has to be analyzed

### **Current summary: Secretory pathway and some lectins**



### **Conclusions**

- The select affinity of VIP36 relative to ERGIC53 for O-glycans has been clearly shown in this *in silico* docking method
- Ca<sup>2+</sup>-dependent binding mimicked and strongly reduced in the the proposed D131N-N166D mutant applies to the core2 O-glycan in particular and a few others, similar to the lectin from *Bauhinia purpurea* an expanded binding site with specific interactions on the perimeter of the central binding cavity can be expected
- Fucosylation by Fut8 may present one signal for enhanced cancer progression with fucosylated N-glycans, the role of core-fucosylation machinery has been demonstrated in melanomas
- Cancer progression has been shown to entail VIP36 as a prognostic marker in renal cancer (Uhlen et al. 2017)